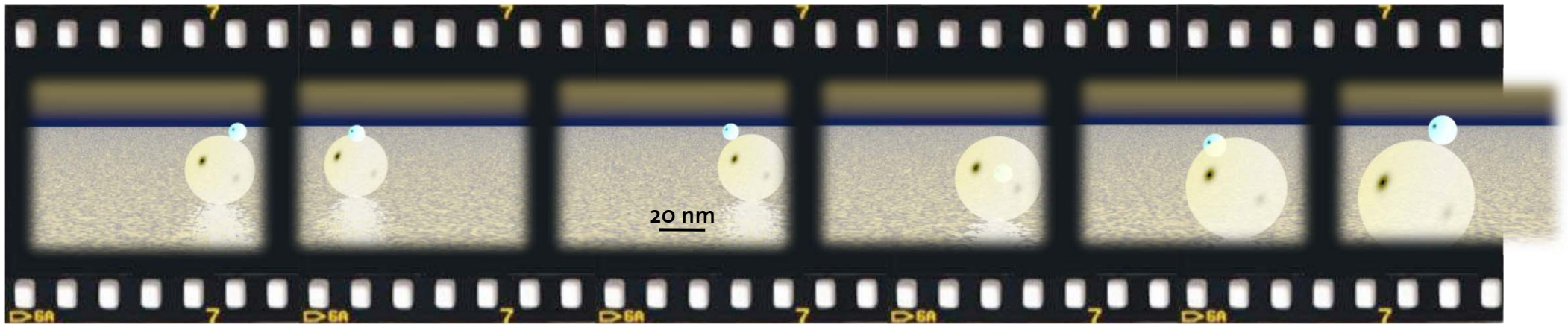


Optical microscopy at the limit



*opt***ETH**
www.opteth.ethz.ch

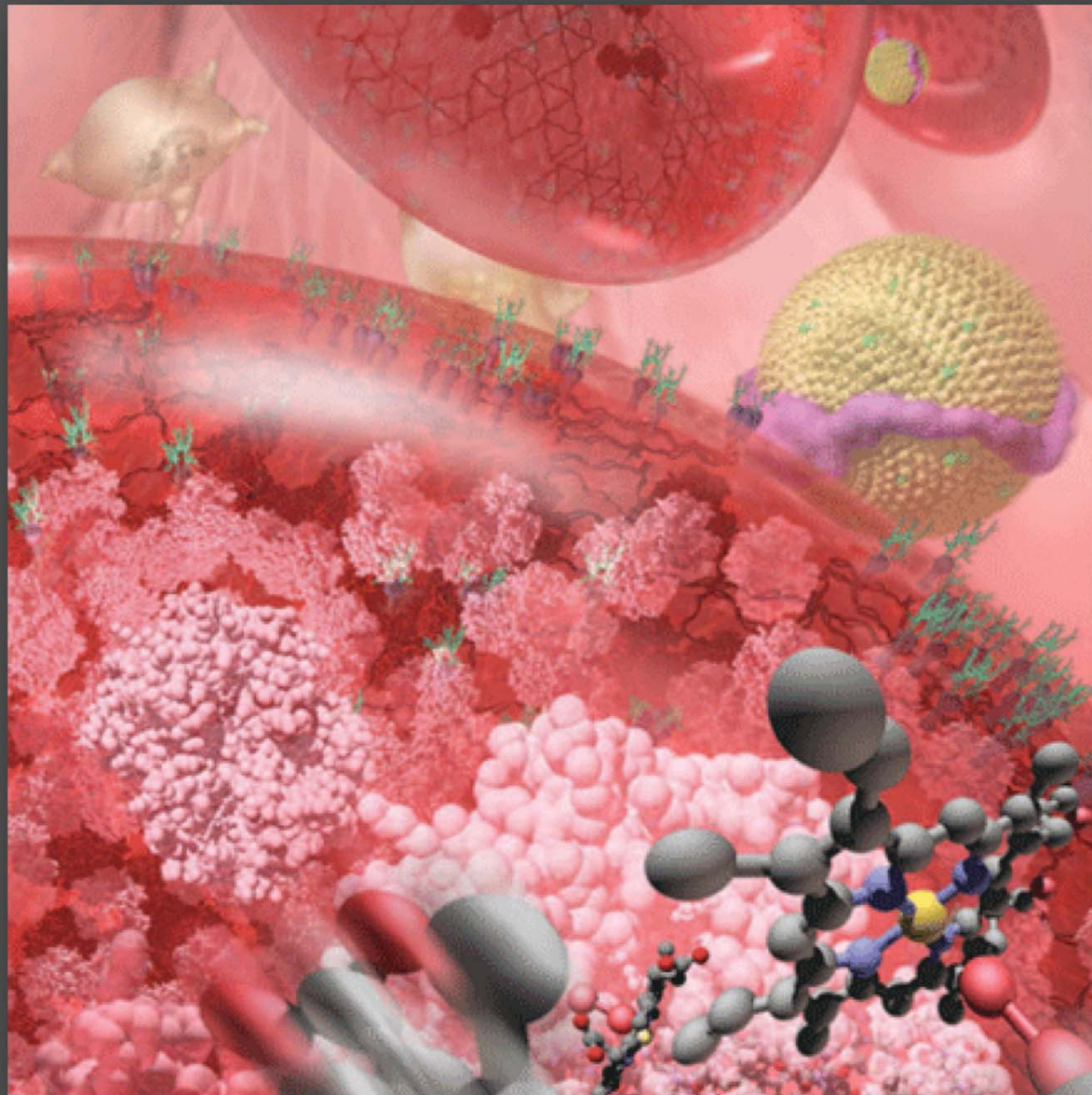


Philipp Kukura,¹ Helge Ewers,² Christian Müller,¹ Alois Renn,¹
Ari Helenius² and Vahid Sandoghdar¹

¹Nano-optics group, Laboratory of Physical Chemistry, ETH Zurich

²Institute of Biochemistry, ETH Zurich

The desire to see small

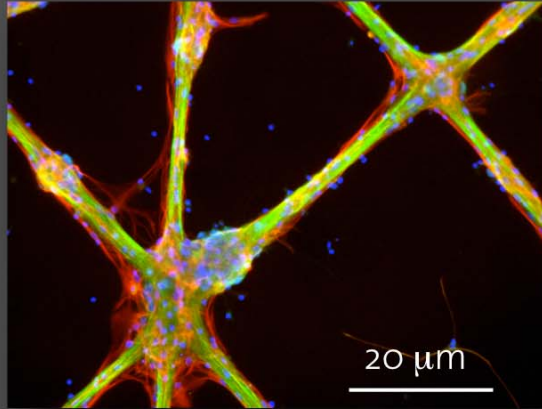


© Linda Nye and the Exploratorium Visualization Laboratory

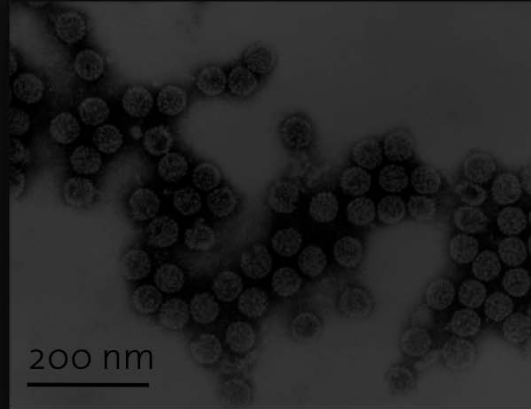
Different ways to see small

Using electromagnetic radiation

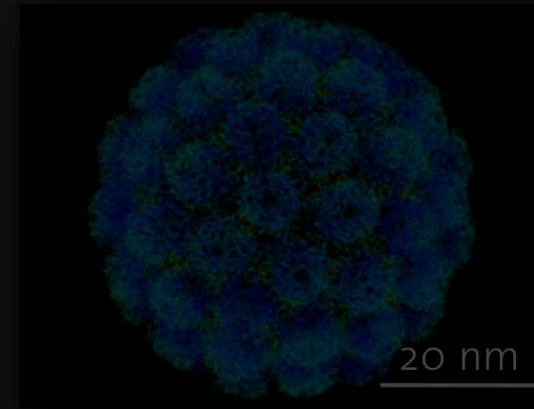
Optical Microscopy



Electron Microscopy

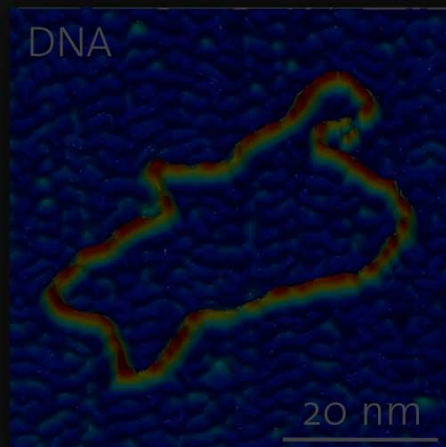


X-Ray Crystallography

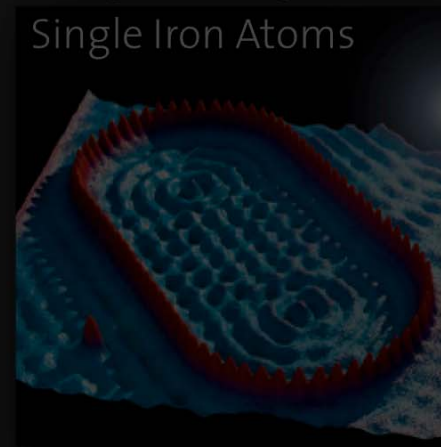


Using a scanning probe

Atomic Force Microscopy

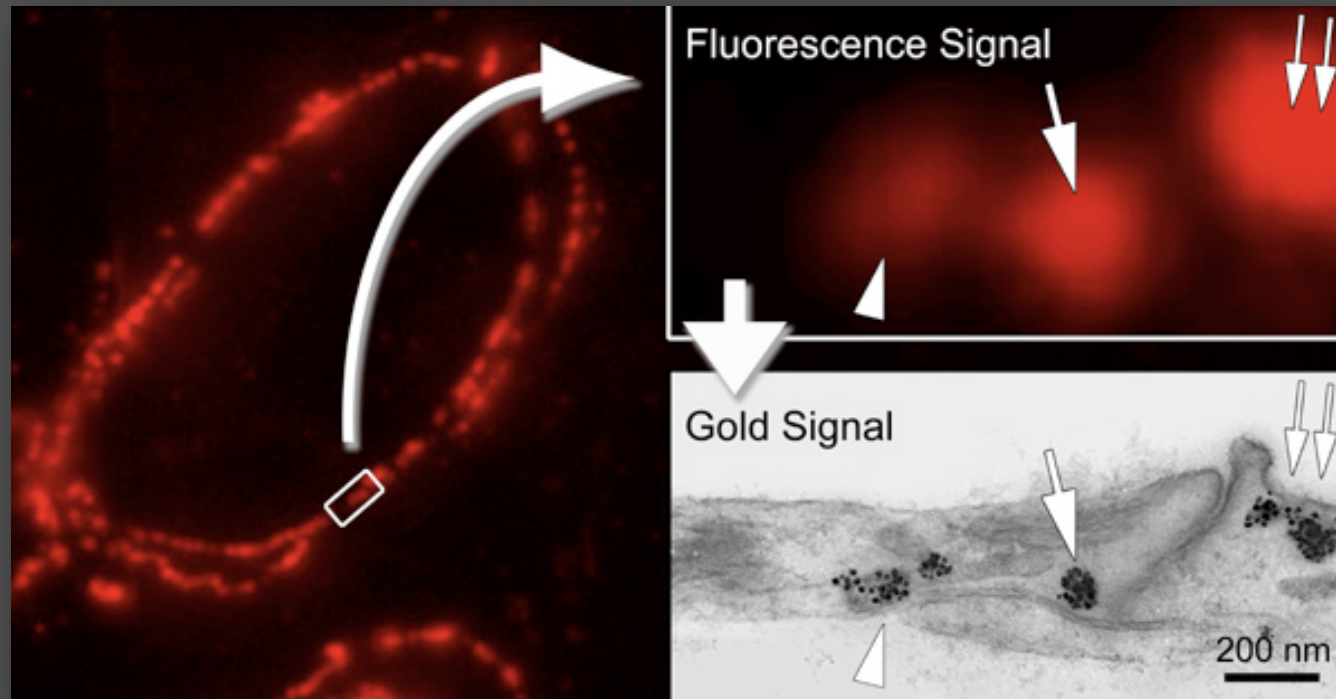
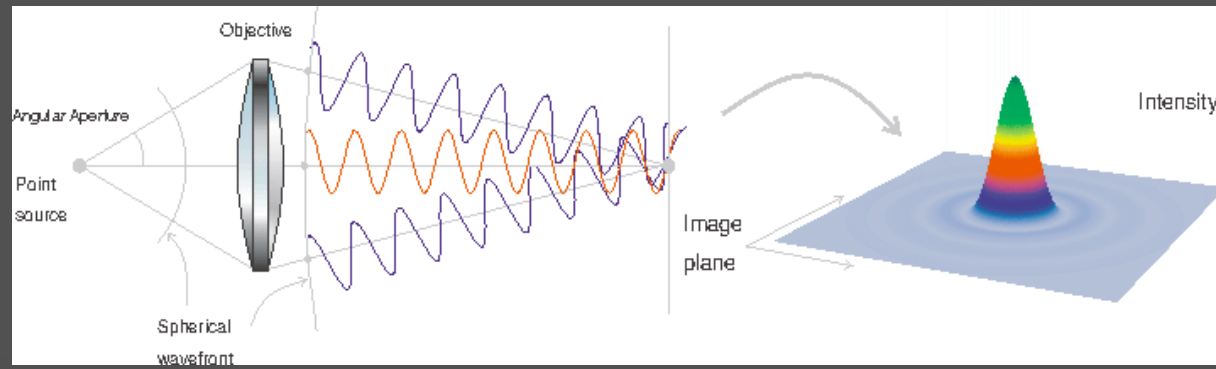


Scanning Tunneling Microscopy



For real time imaging of live specimens – optical microscopy wins

One big problem: the diffraction limit



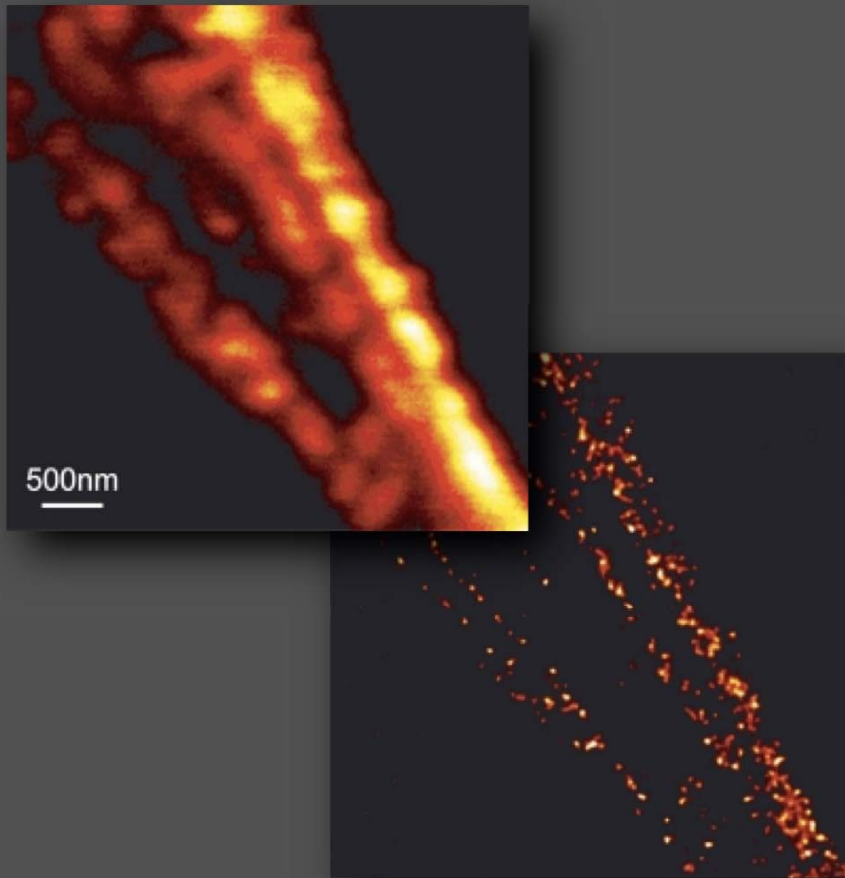
www.nanoprobes.com

No matter how small the light source – it looks (at least) as big as $\lambda/2$: > 200 nm

Recent developments to get around it

STED

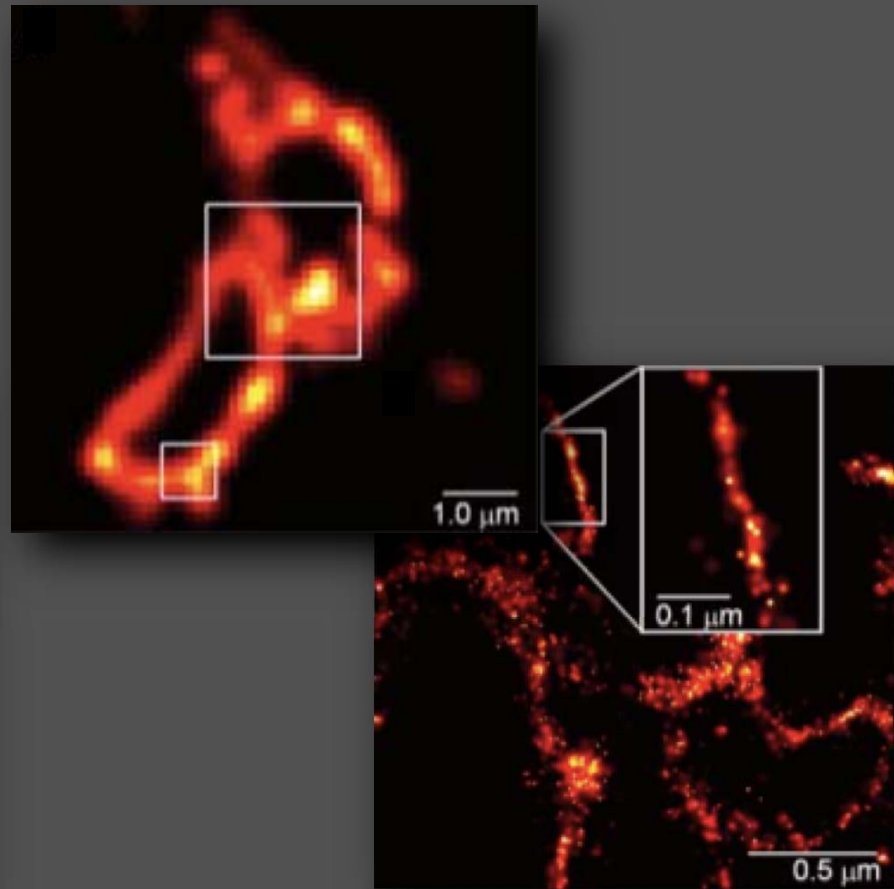
S. W. Hell, *Science* **316**, 1153 (2007)



Nonlinear contrast improvement

PALM, STORM, etc.

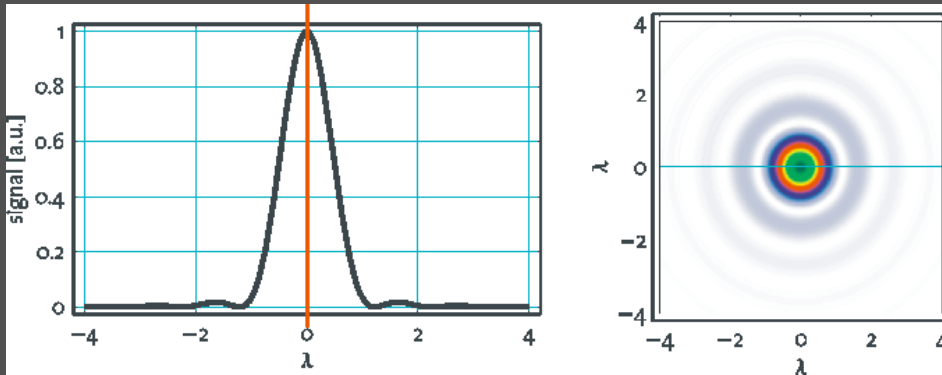
E. Betzig et al., *Science* **313**, 1643 (2006)
M. J. Rust et al., *Nat. Methods* **3**, 793 (2006)
S. Hess et al., *Biophys. J.* **91**, 4258 (2006)



Single molecule localization

Single molecule localization

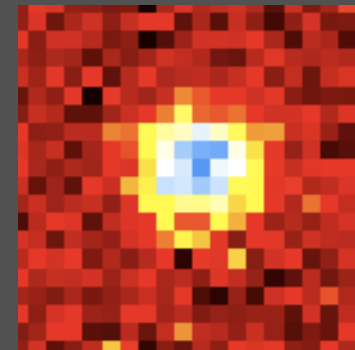
Determine center through statistical fit of ideal point spread function



Accuracy scales with the number of detected photons, N :

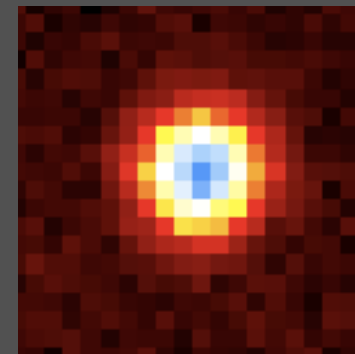
$$N^{-1/2}$$

$$N = 100$$

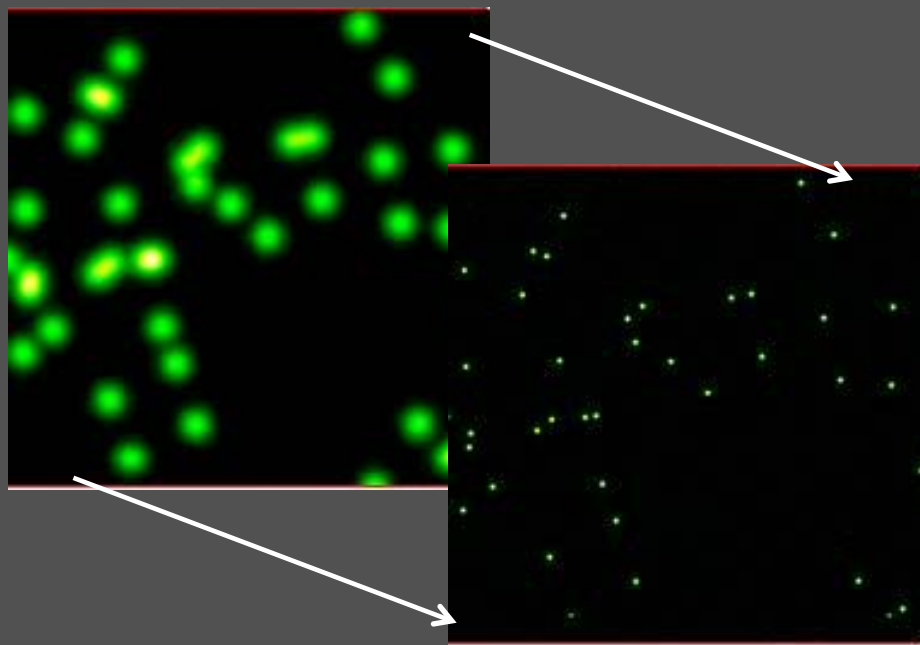


$$\sigma = \sim 10 \text{ nm}$$

$$N = 10000$$

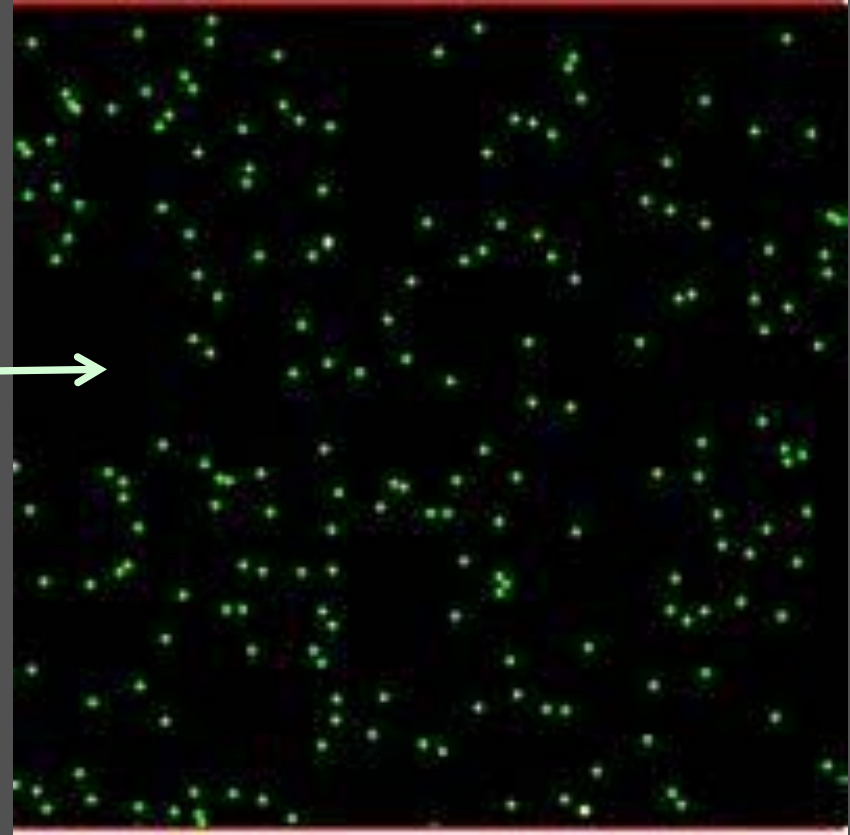
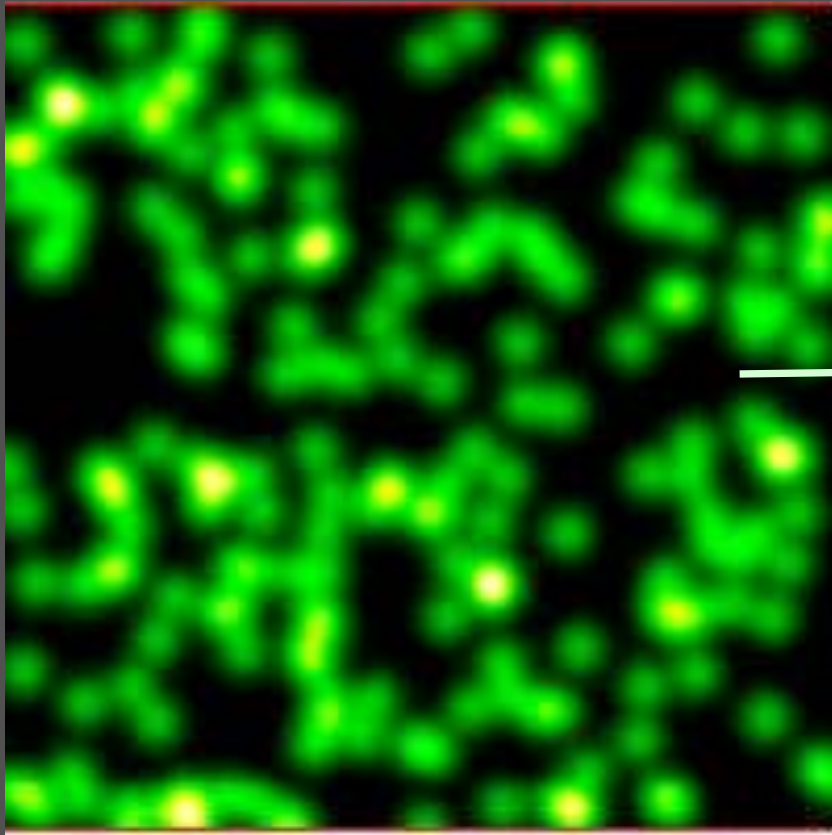


$$\sigma = \sim 1 \text{ nm}$$

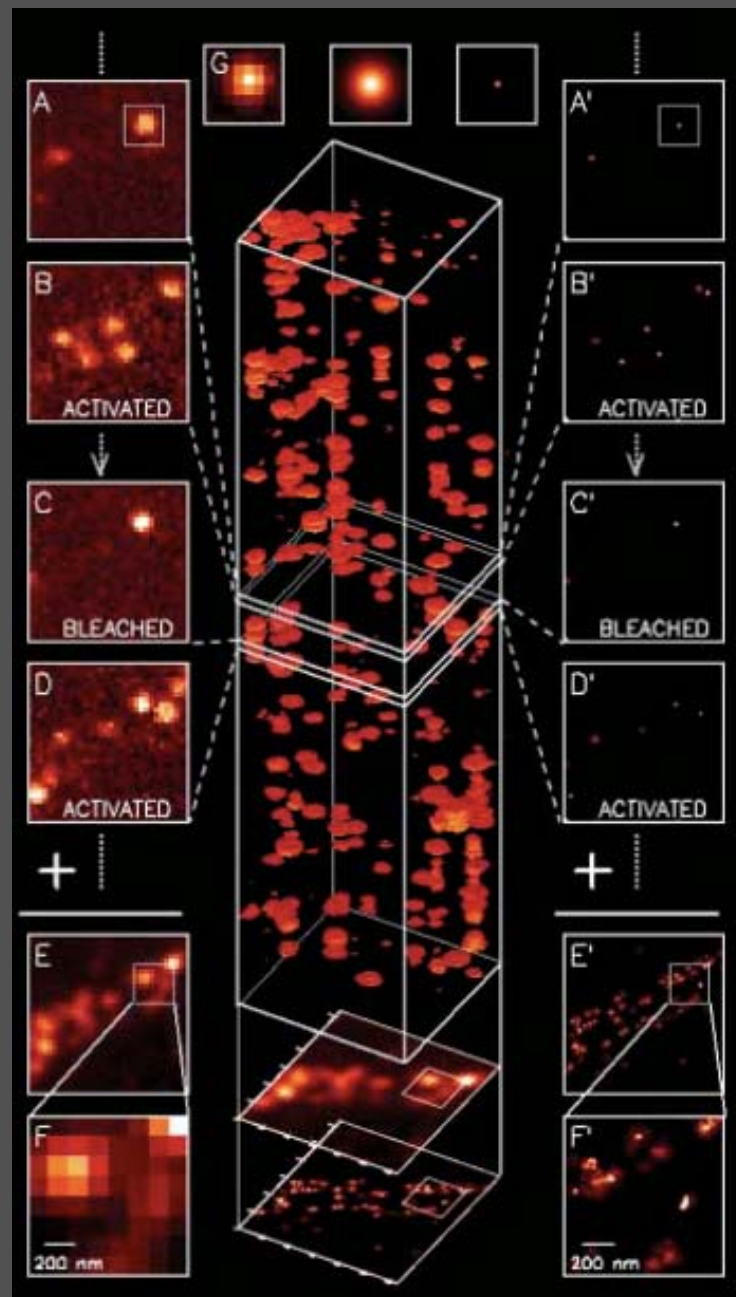


The trouble with *colocalization*

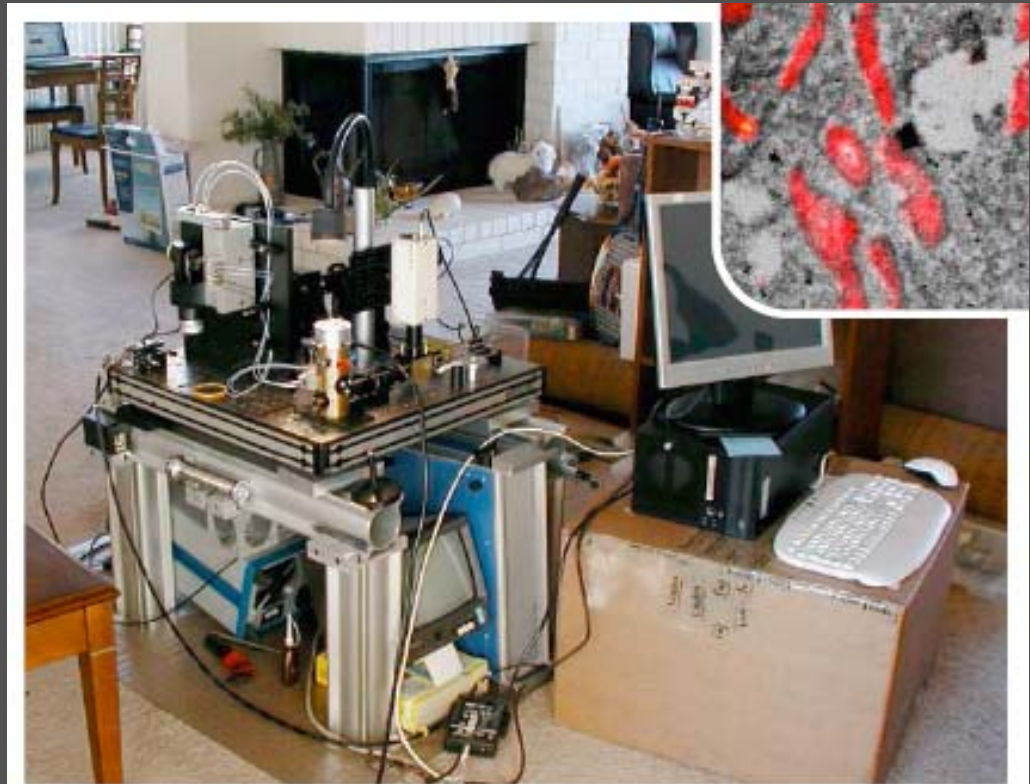
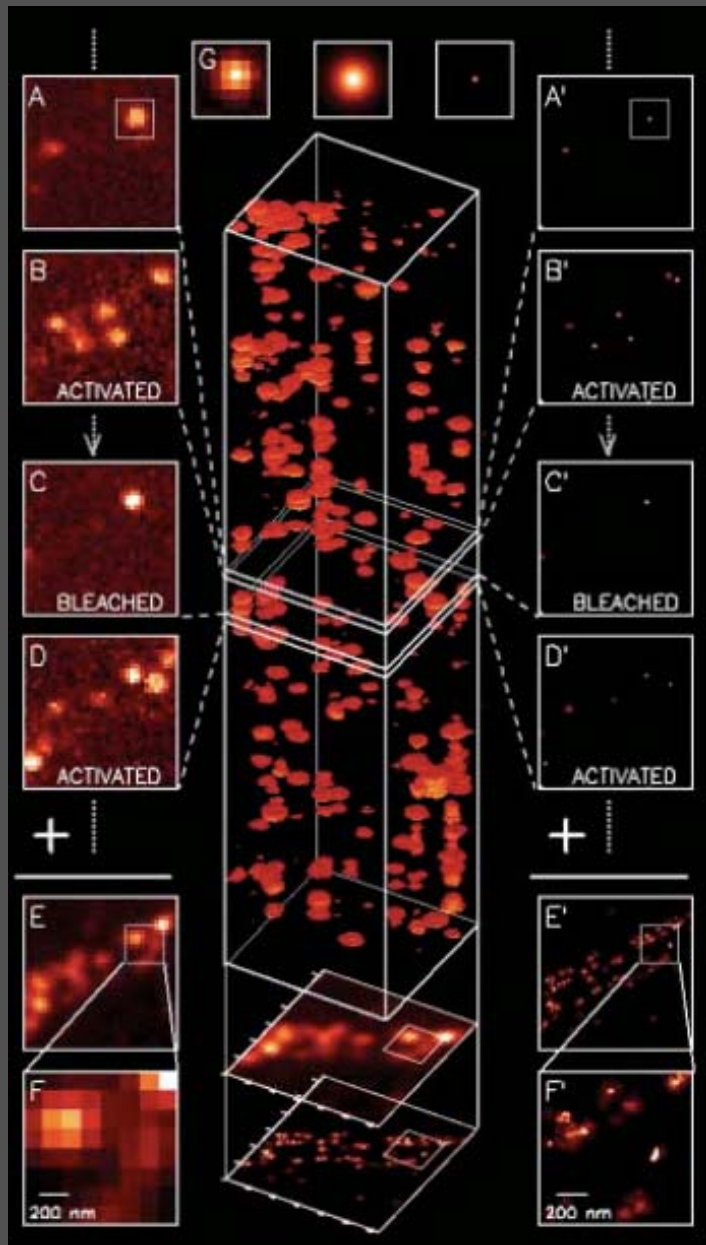
What to do if the density becomes too high?



Photoactivation

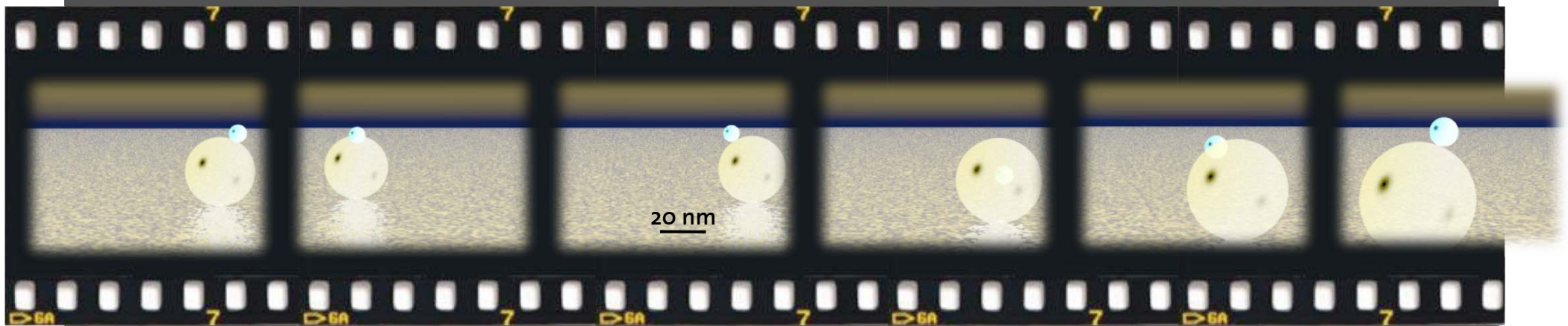


Photoactivation



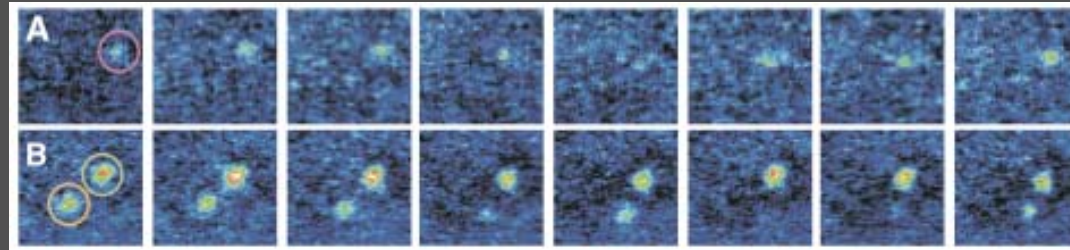
Up close. A high-tech microscope, assembled in a living room (*above*), revealed molecules (red, *inset*) nanometers apart inside a cell's mitochondria.

What about dynamics?



Tracking single molecules

Visualizing the infection pathway of an adeno-associated virus

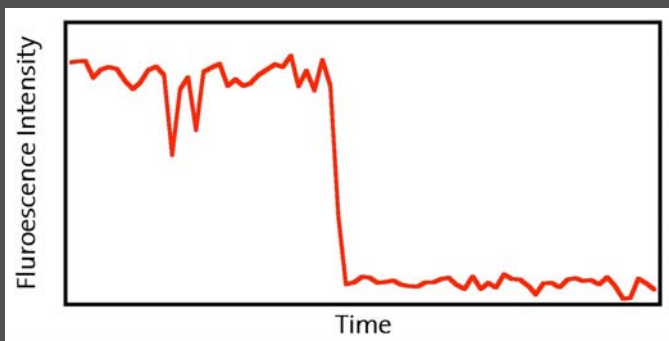
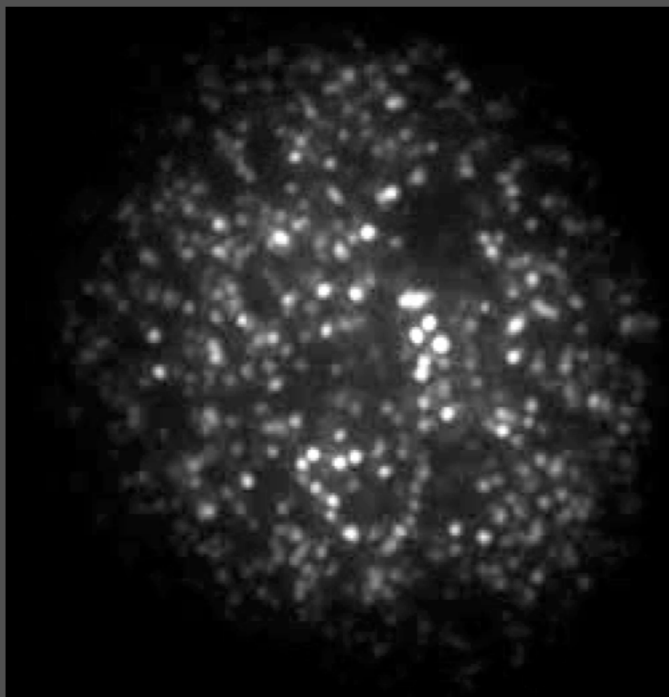


Localize virus in each frame
Connect positions to create trajectories

G. Seisenberger et al., *Science* **294**, 1929 (2001)

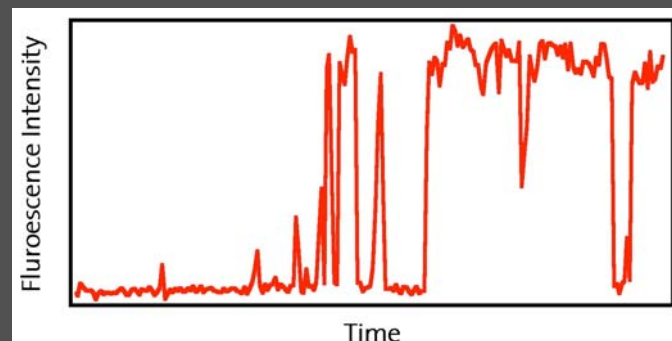
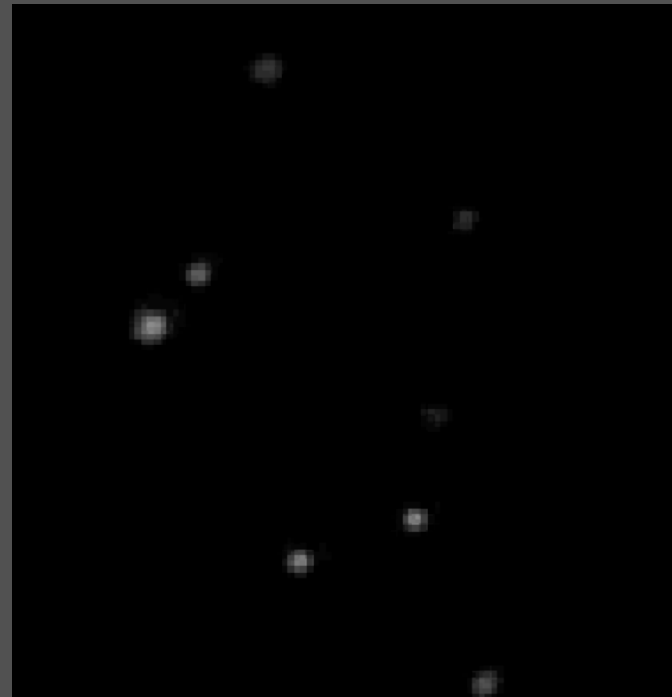
Single emitters: light sources with ticks

Single Cy3 molecules



„Bleaching“

Single Quantum Dots



„Blinking“

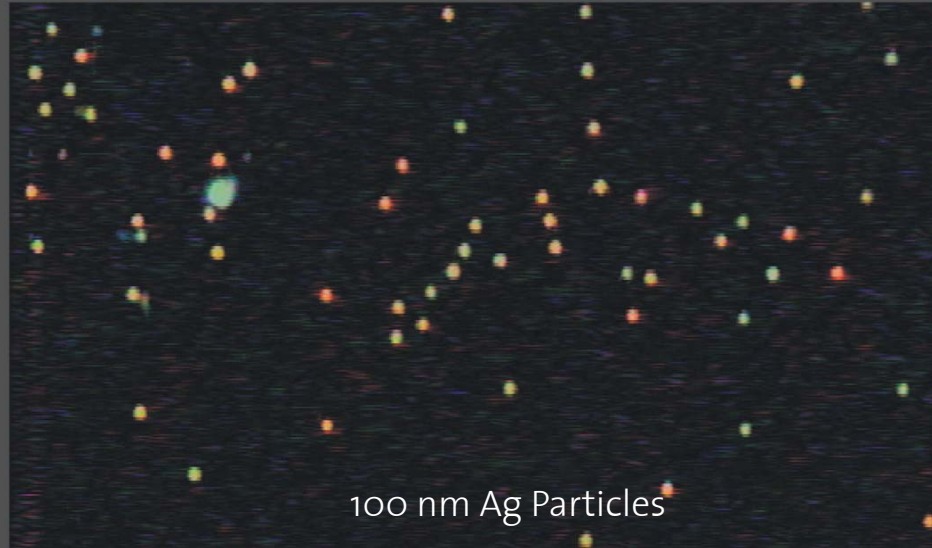
An alternative: light scattering



Milk does not blink or bleach!

Scattering detection of tiny particles

via scattering intensity: need to eliminate background scattering
→ dark-field illumination, total internal reflection



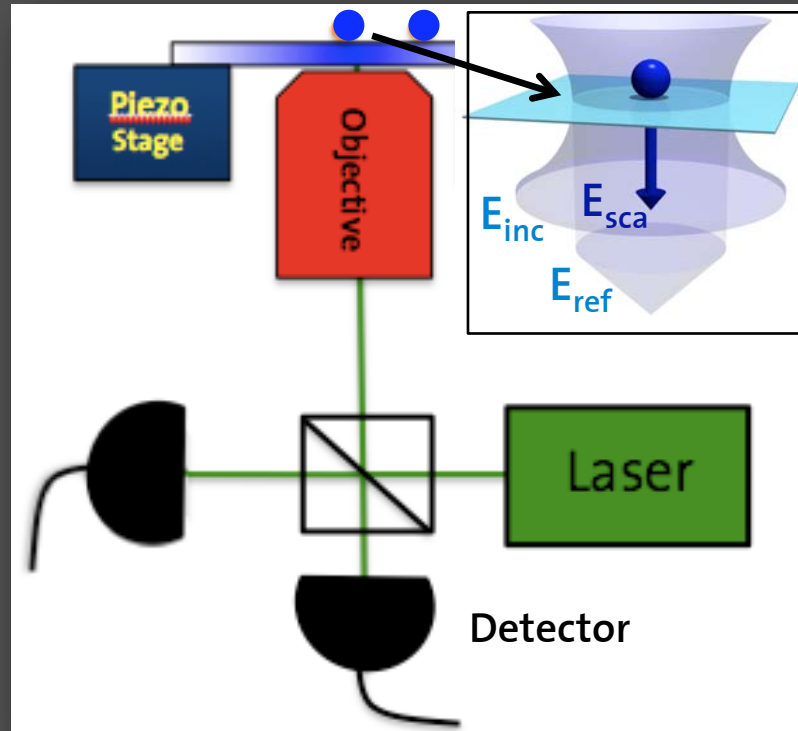
scattering amplitude, $s(\lambda)$

$$s(\lambda) = \eta\alpha(\lambda) = \eta\varepsilon_{med}(\lambda) \frac{\pi D^3}{2} \frac{\varepsilon_{part}(\lambda) - \varepsilon_{med}(\lambda)}{\varepsilon_{part}(\lambda) + 2\varepsilon_{med}(\lambda)}$$

The scattering cross section scales as D^6 thus rapidly drops below the noise
Detection limit: ~30 nm Au

interferometric scattering detection: iSCAT

Simplified experimental setup



Scattered field:

$$E_{\text{ref}} \sim r e^{-i\pi/2} E_{\text{inc}}$$

Reflected field:

$$E_{\text{sca}} \sim |\alpha| e^{-i\phi} E_{\text{inc}}$$

$$I_{\text{det}} = |E_{\text{ref}}|^2 + |E_{\text{sca}}|^2 + 2\text{Re}\{E_{\text{ref}}^* E_{\text{sca}}\}$$

$$I_{\text{det}} = I_{\text{ref}} + I_{\text{sca}} + I_{\text{int}}$$

Scattering contrast:

$$S = \frac{I_{\text{int}}}{I_{\text{ref}}}$$

Background:

$$B = \frac{1}{N^{-(1/2)}}$$

Signal-to-noise ratio:

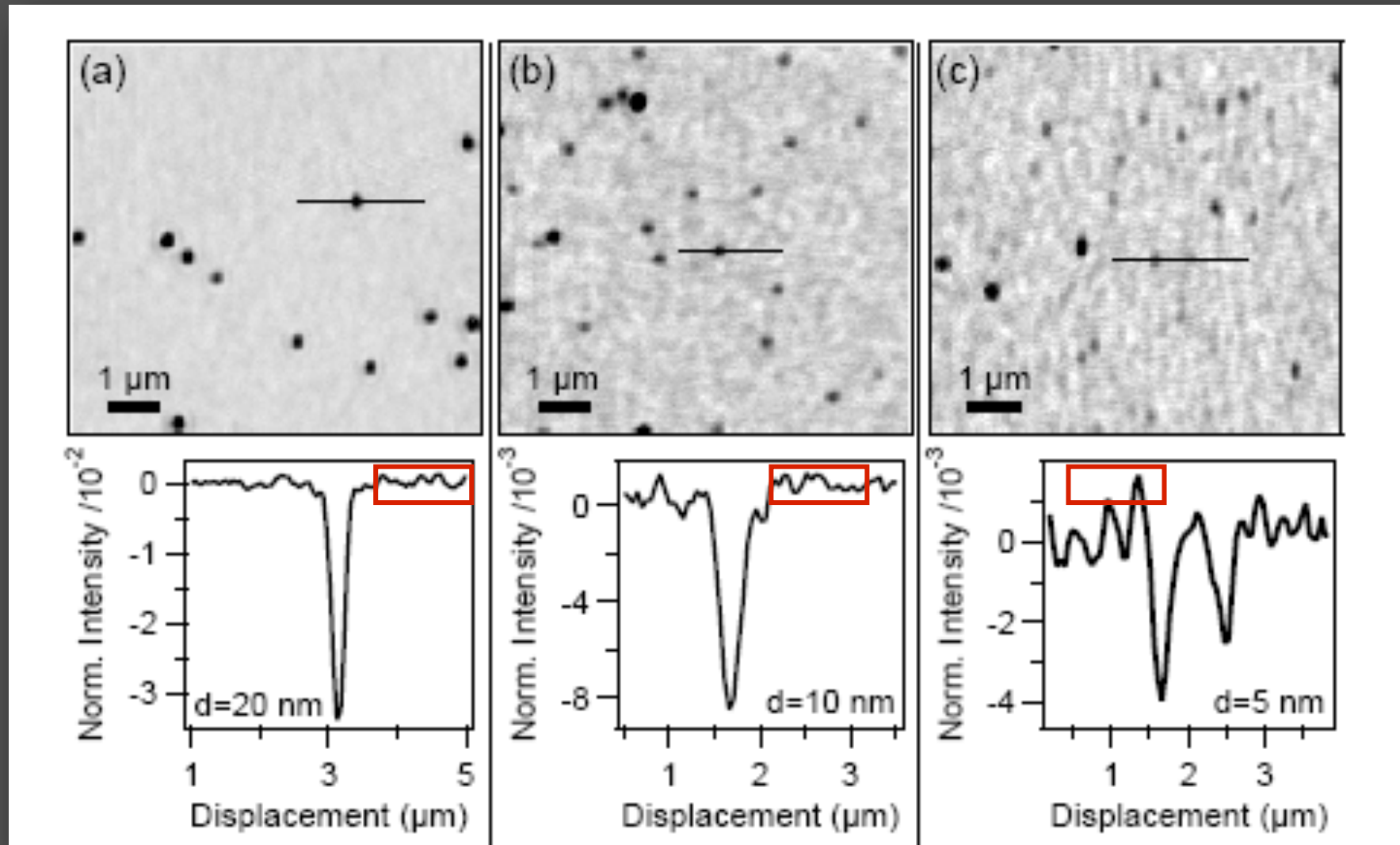
$$\text{SNR} = \frac{S}{B} = S \times N^{(1/2)}$$

Interferometric detection much less sensitive to particle size: D^3 vs D^6

SNR scales with the number of incident photons

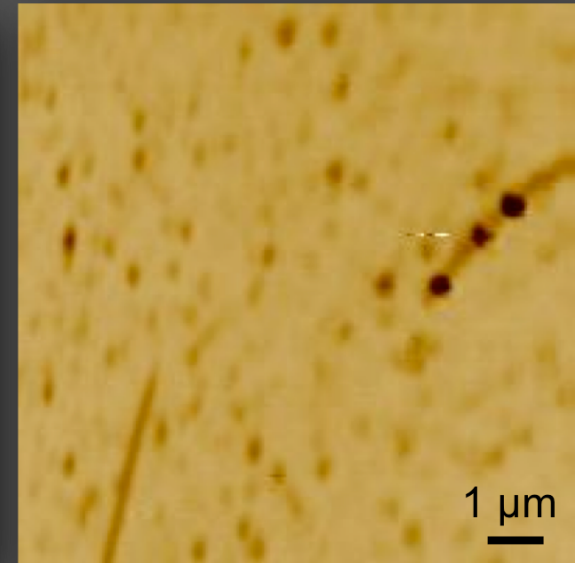
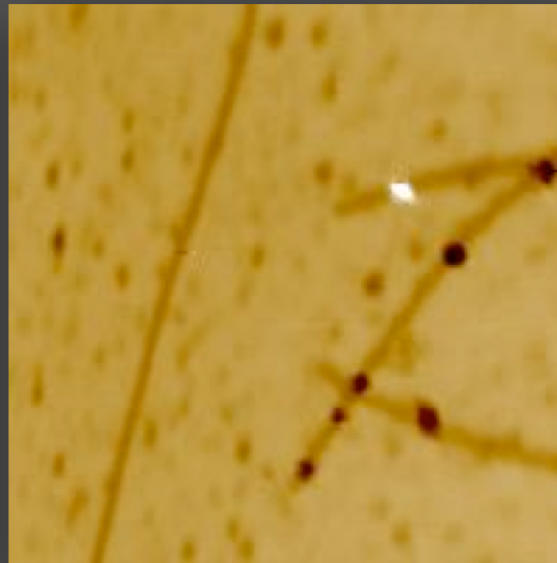
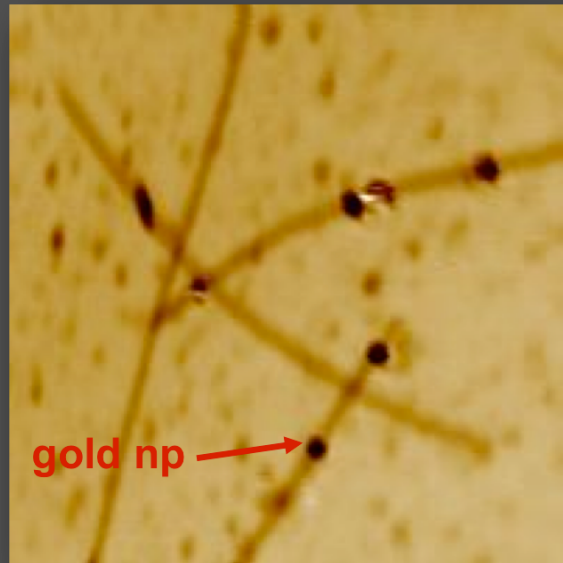
Detection of *nearly* molecular sized scatterers

Piezo raster scan of gold nanoparticles on glass in water



Goldnanoparticles as biological labels

Microtubules labeled with 40 nm gold particles

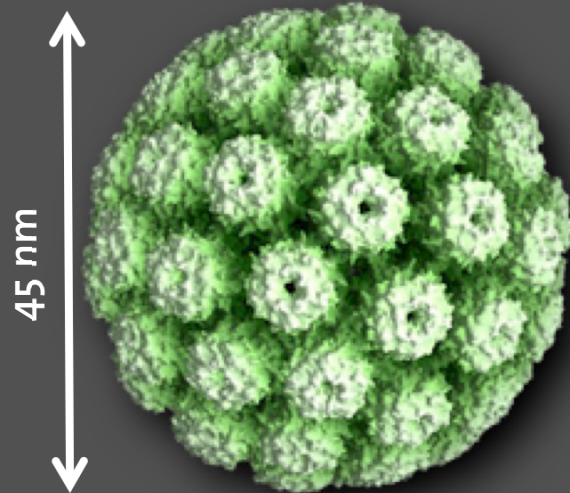


$$s(\lambda) = \eta\alpha(\lambda) = \eta\varepsilon_{med}(\lambda) \frac{\pi D^3}{2} \frac{\varepsilon_{part}(\lambda) - \varepsilon_{med}(\lambda)}{\varepsilon_{part}(\lambda) + 2\varepsilon_{med}(\lambda)}$$

Biological material also produces a signal since $\varepsilon_{part}(\lambda) \neq \varepsilon_{med}(\lambda)$

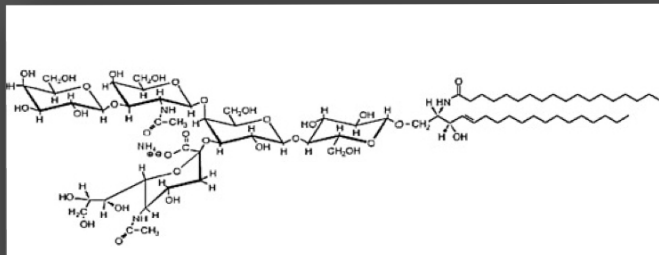
Label-free detection of single viruses

Simian virus 40

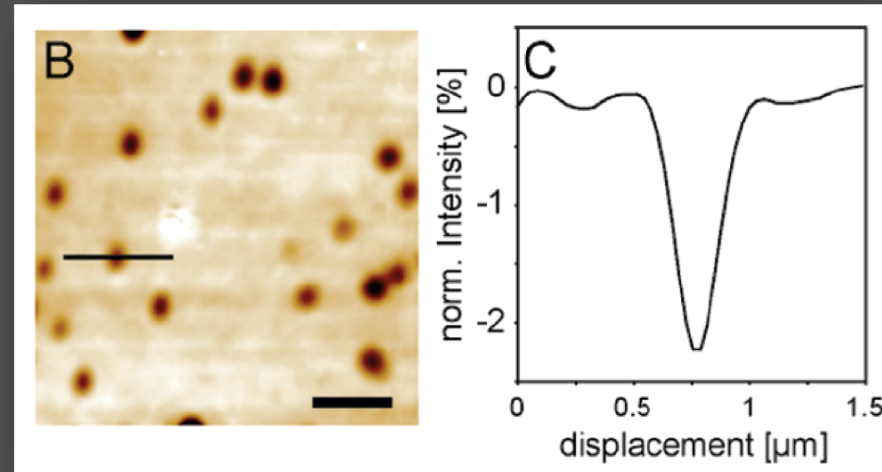


Liddington et al., *Nature* **354**, 6351 (1991)

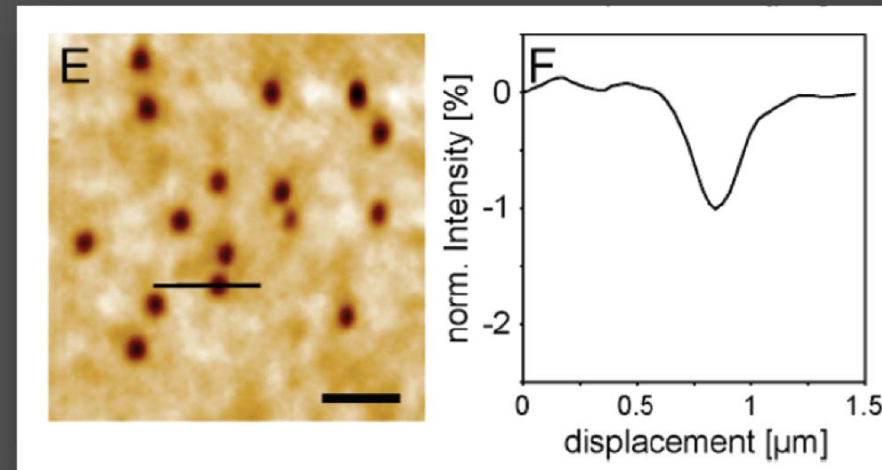
- Non-enveloped DNA tumor virus
- 72 pentamers of viral protein 1
- Cellular receptor: Glycol moiety of GM1



Single viruses bound to cover glass



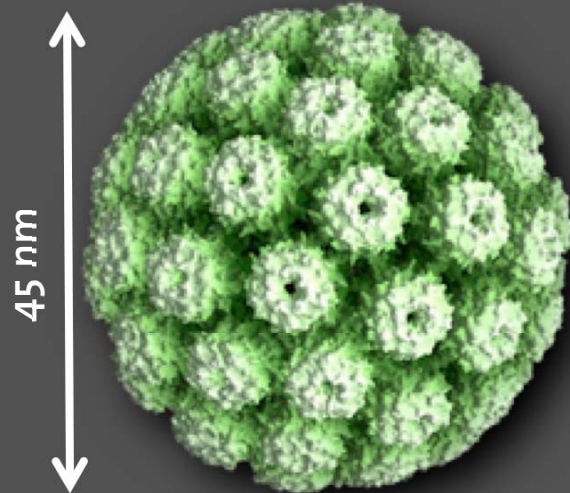
Virus like particles (empty protein shell)



H. Ewers, V. Jacobsen, E. Klotzsch, A. E. Smith, A. Helenius, V. Sandoghdar, *Nano Lett.* **7**, 2263 (2007).

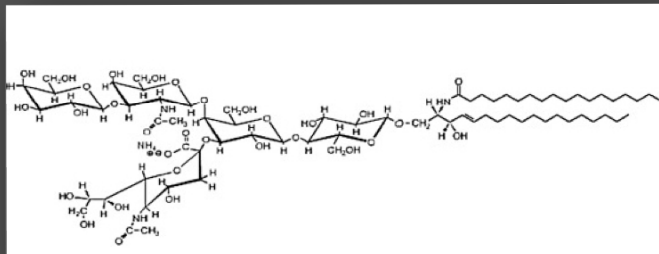
Label-free detection of single viruses

Simian virus 40

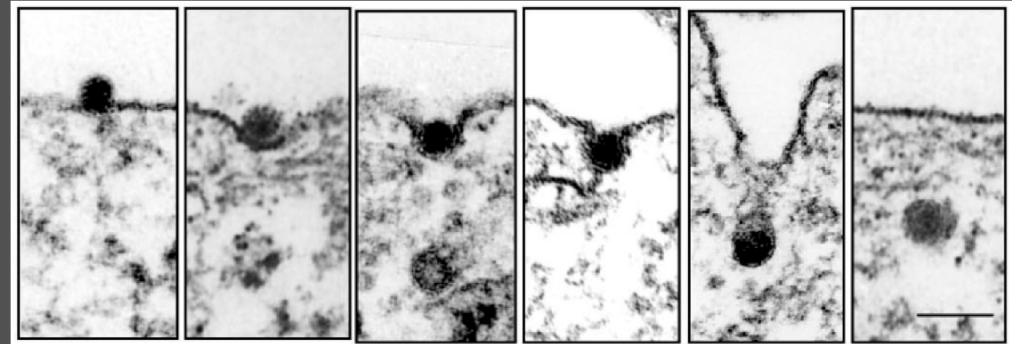


Liddington et al., *Nature* **354**, 6351 (1991)

- Non-enveloped DNA tumor virus
- 72 pentamers of viral protein 1
- Cellular receptor: Glycol moiety of GM1



SV40 endocytosis



E. M. Damm, L. Pelkmans, J. Kartenbeck, A. Mezzacasa, T. Kurzckalia and A. Helenius, *J. Cell Biol.* **168**, 477 (2005)



M. J. Lehmann, N. M. Sherer, C. B. Marks, M. Pypaert and W. Mothes, *J. Cell Biol.* **170**, 317 (2005)

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